

For Veterinary Use Only

READ ALL INSTRUCTIONS BEFORE BEGINNING THE TEST

 RIDX™ CPV Ag Test Kit

[Catalogue Number: CGM-CPG-11, CGM-CPG-12]

Introduction

Canine parvovirus (CPV), a nonenveloped single-stranded DNA (ssDNA) virus (family Parvoviridae, genus *Protoparvovirus*), is a highly successful pathogen that has sustained pandemic circulation in dogs for more than 40 years¹. Canine parvoviruses are in the form of CPV-1 and CPV-2. Most dogs of CPV-1 infection are asymptomatic². CPV-2 causes serious gastrointestinal and respiratory disease and has variants of CPV-2a, CPV-2b, and CPV-2c³.

CPV enters through the mouth from the ground or floor and its incubation period is three to seven days before dogs appear symptoms⁴. CPV infected dogs appear lethargy, loss of appetite, abdominal bloating and pain, fever, or low body temperature (hypothermia), vomiting, and severe, often bloody, diarrhea⁵.

Principle

The RIDX™ CPV Ag Test Kit is a lateral flow chromatographic immunoassay for the qualitative detection of CPV antigens in canine feces. This kit shows two letters which are the test (T) line and the control (C) line on the surface of the device. If the CPV antigen exists in the sample, it binds to the gold-conjugated CPV antibody. The antigen-antibody complex moves through the membrane by capillary force and responds to the CPV antibody on the test line, resulting in a red line. The control line indicates that the test is performed correctly and should appear when the test is complete.

The highly selective and sensitive monoclonal antibody to CPV is used as a capture and detector in the kit. The RIDX™ CPV Ag Test Kit can detect CPV antigen in canine feces with high accuracy.

Performance

1. Sensitivity & Specificity

		PCR		
		+	-	Total
RIDX™	+	101	0	101
CPV Ag	-	2	214	216
Test	Total	103	214	317

Sensitivity: 98.06% (101/103, *95% CI: 93.19% ~ 99.47%)

Specificity: 100% (214/214, 95% CI: 98.24% ~ 100%)

Diagnostic Agreement: 99.37% (315/317, 95% CI: 97.73% ~ 99.83%)

* 95% CI: 95% Confidence Interval

2. Limit of Detection: 5×10^3 TCID₅₀/mL

3. CPV-2, CPV-2a, CPV-2b diagnosis are also available.

4. Cross-Reactivity

Potentially cross-reactive substances listed below have no effect on the performance of the RIDX™ CPV Ag Test Kit.

Pathogen	Titer	Result
Canine coronavirus	1.00×10^6 TCID ₅₀ /mL	Negative
Canine distemper virus	1.00×10^5 TCID ₅₀ /mL	Negative
Canine influenza virus	1.00×10^6 EID ₅₀ /mL	Negative
<i>Escherichia coli</i>	3.56×10^8 CFU/mL	Negative
<i>Giardia</i> spp.	1.42×10^5 Cysts/µL	Negative
<i>Salmonella</i> spp.	1.00×10^6 CFU/mL	Negative

Kit Components

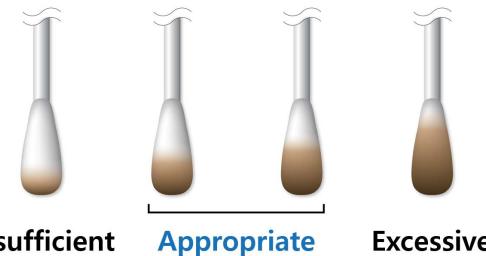
Component	Quantity/kit by CAT No.	
	CGM-CPG-11	CGM-CPG-12
1 CPV Ag test device	10	2
2 Sample dilution buffer	10	2
3 Disposable swab	10	2
4 Disposable dropper	10	2
5 Instructions for use	1	1

Storage & Stability

1. Store the test kit at 2~30°C (35.6~86.0°F). **Do not freeze.**
2. Do not store the test kit in direct sunlight.
3. The test kit is stable within the expiration date marked on the package label.

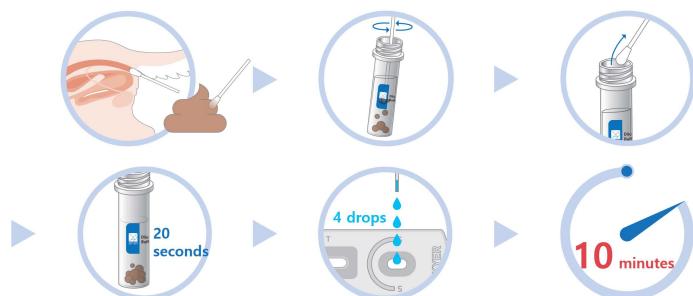
Sample Preparation

1. **Canine fecal swab** should be used for this test.
2. The samples should be tested immediately after collection.
3. If samples cannot be tested immediately, they should be stored at 2~8°C (35.6~46.4°F) for up to 24 hours. For longer storage, freeze at -20°C (-4°F) or below. But, results from samples frozen for over one month may differ from those obtained before freezing. Frozen samples should be brought to room temperature (15~30°C/59~86°F) before use.
4. The amount of fecal sample with swab may affect the results. It is required to follow the swab amount of feces as shown in the picture below. The excessive fecal amount may induce a false positive result and slow migration.



◆ Test Procedure

1. All reagents and samples must be at room temperature (15~30°C /59~86°F) before use.
2. Collect fecal samples using a swab.
3. Put the swab into the sample dilution buffer and stir the solution with the swab to disperse the sample into the buffer (approximately 10 seconds).
4. Remove the swab from the sample dilution buffer.
5. Wait for 20 seconds to settle down the large particles.
6. Remove the test device from the pouch and place it on a flat and dry surface.
7. Take the supernatant sample in the tube by using a disposable dropper.
8. Apply 4 drops of the mixed sample into the sample hole (S), drop by drop vertically.
9. Read test result at 10 minutes. **Do not read results that appear after 10 minutes.**



[Summary of Test Procedure]

◆ Interpretation of Results

1. Positive result

Test (T) line and control (C) line within the result window indicate the presence of CPV antigen.



2. Negative result

Only control (C) line appears in the result window.



3. Invalid results

If the control (C) line does not appear, the result might be considered invalid. The sample should be retested.



◆ Precautions

1. This test kit is for veterinary *in vitro* diagnostic use only for dogs. Do not use this test kit for other animals.
2. This rapid kit is only for preliminary screening. The final decision should be made by a qualified veterinarian based on the results of this kit, clinical symptoms and evaluation by a veterinarian, and, if necessary, the results of additional detailed diagnostic procedures.
3. The test device is sensitive to humidity and heat. Use the test device within 10 minutes after removing the foil pouch.
4. Do not touch the membrane of the test device.

5. The device should not be used if the foil pouch is damaged or has been stored with the seal open.
5. Do not use an expired test kit. The expiration date is marked on the package label.
7. Do not reuse the components (device, buffer, dropper, and swab).
8. Do not mix components from different lot numbers because the components in this kit have been quality control tested as a standard batch unit.
9. Decontaminate and dispose of all samples, used kits, and potentially contaminated materials in the accordance with national and local regulations.
10. All samples should be handled as being potentially infectious. Wear protective gloves while handling samples. Wash hands thoroughly afterward.

◆ References

1. Voorhees IEH, Lee H, Allison AB, Lopez-Astacio R, Goodman LB, Oyesola OO, Omobowale O, Fagbohun O, Dubovi E, Hafenstein S, Holmes EC, Parrish CR. Limited Intrahost Diversity and Background Evolution Accompany 40 Years of Canine Parvovirus Host Adaptation and Spread. *J Virol.* 2020; 94(1): 1162-19.
2. Goddard A, Leisewitz AL. Canine Parvovirus. Veterinary Clinics of North America. *Small Animal Practice* 2010; 40(6): 1041-1053.
3. Cavalli A, Martella V, Desario C, Camero M, Bellacicco AL, Palo P, Decaro N, Elia G, Buonavoglia C. Evaluation of the Antigenic Relationships among Canine Parvovirus Type 2 Variants. *Clin Vaccine Immunol.* 2008; 15(3): 534-539.
4. Nandi S, Kumar M. Canine Parvovirus: Current Perspective Indian. *J Virol.* 2010; 21(1): 31-44.
5. Meunier PC, Cooper BJ, Appel MJ, Slauson DO. Pathogenesis of canine parvovirus enteritis: the importance of viremia. *Vet Pathol.* 1985; 22(1): 60-71.

◆ Symbol Descriptions

	License number
	Catalogue number
	Batch code, Lot number
	Consult instructions for use
	Contains sufficient for $\langle n \rangle$ tests
	Do not reuse
	<i>In vitro</i> diagnostic medical device
	Temperature limitation
	Do not use, if the package is damaged
	Upper side
	Manufacturer



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